

Trifluoroacetyl Neonicotinoid Insecticides with Enhanced Hydrophobicity and Effectiveness

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Neonicotinoids with nitro- or cyanoimino substituents are extensively utilized as plant-mobile (systemic) insecticides controlling the piercing–sucking insect pests. This investigation considers structural features of neonicotinoids with trifluoroacetyl pharmacophores, which may confer enhanced hydrophobicity and effectiveness. Fifteen trifluoroacetyl neonicotinoid analogues [=NC(O)-CF₃ and =CHC(O)CF₃] are therefore prepared to evaluate the hydrophobicity index, toxicity to houseflies (*Musca domestica*), and binding affinity to the *Musca* nicotinic receptor. The =NC(O)CF₃ and =CHC(O)CF₃ compounds showed a higher hydrophobicity than that of nitro- or cyanoimino analogues. The intrinsic insecticidal activities (defined by intrathoracic injection with a synergist pretreatment) of test compounds were well-correlated to their target site potencies. Although nitro or cyano neonicotinoids were not toxic via the topical application route in the absence of a synergist, trifluoroacetyl analogues exhibited excellent insecticidal activity under the same condition. Accordingly, the increased hydrophobicity of trifluoroacetyl neonicotinoids presumably improves the penetrability of compound into insect integument and insecticidal effectiveness.

KEYWORDS: Neonicotinoid insecticides; nicotinic acetylcholine receptor; trifluoroacetyl pharmacophore

INTRODUCTION

Neonicotinoids, exemplified by imidacloprid (IMI) and thiacloprid (THIA) providing nitro- and cyanoimino pharmacophores, respectively (**Figure 1a**), are utilized throughout the world as systemic (plant-mobile) insecticides for crop protection against piercing-sucking insect pests (I-5). Neonicotinoid selectively acts on the insect nicotinic acetylcholine receptor (nAChR), and the binding site interactions in chemical or atomic resolution have been defined by comparative chemical and structural neurobiology approaches using mollusk acetylcholine binding protein (AChBP), which is a suitable structural surrogate of the extracellular ligand binding domain of the nAChRs (6-10).

In our previous studies, the systematic pharmacophore modifications of neonicotinoid molecules gave highly active compounds with diverse chemotypes of pharmacophores, and three-dimensional structural models of the binding site interactions were predictive of the observed structure—activity relation-ships (SARs) (11-13). Intriguingly, a neonicotinoid analogue with a trifluoroacetyl substituent, replacing the nitro or cyano pharmacophore, is highly insecticidal, apparently associated with the unique binding site interactions of the trifluoroacetyl moiety (**Figure 1b**) (11, 14). The present investigation examines the hypothesis that the trifluoroacetyl substituent enhances the

hydrophobicity of neonicotinoid and thereby confers the improved insecticidal effectiveness relative to those of the nitro and cyano compounds. Therefore, we designed and prepared trifluoroacetyl neonicotinoid analogues (**Figure 2**) to comprehensively consider the SARs regarding the nAChR affinity and insecticidal potency in comparison with those of the nitro and cyano neonicotinoids.

MATERIALS AND METHODS

Chemistry. Compounds 1–5, 8, 10, 15, and 17–19 were available from our previous studies or prepared according to published methods (*12*, *14–18*). Synthesis procedures for the new chemicals (compounds 6, 7, 9, 11–14, 16, and 20–22) are summarized in Scheme 1, and the details and analytical results for structural confirmation are given in the Supporting Information. All melting points (mp) are uncorrected. ¹H, ¹³C, and ¹⁹F NMR spectra were recorded using a JEOL ECA-500 spectrometer at 500, 125, and 376.3 MHz, respectively. The chemical shifts were recorded in δ (ppm), and the coupling constants J_{H-H} were recorded in Hz unless otherwise stated. Mass spectra were recorded at 70 eV with the JEOL JMS-700 instrument.

Physicochemistry. Partition coefficients of test compounds between 1-octanol and water (log P_{ow}) were determined by Kagabu et al. (18). The maximal absorption wavelength (λ_{max}) of neonicotinoids was measured with a HITACHI U-4000S spectrophotometer.

Biology. The potency of test compounds as inhibitors of $[{}^{3}H]IMI$ binding to the native housefly (*Musca domestica*) brain nAChR was assayed according to Tomizawa et al. (19). IC₅₀ values (molar concentrations of test compounds necessary for 50% inhibition of specific $[{}^{3}H]IMI$ binding) were determined by iterative nonlinear least-squares regression

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trifluoroacetylimino analogue

Figure 1. (a) Structures of representative neonicotinoid insecticides IMI and THIA. (b) Predictive binding site interactions of a trifluoroacetyl neonicotinoid (compound 15) with the agonist binding pocket of the *Aplysia californica* AChBP (*11*), which is a structural surrogate for the insect nAChR extracellular domain (*7*, *9*). A water molecule near the pyridine nitrogen atom, captured in the AChBP-IMI crystal structure (Protein Data Bank code 3C79) (*8*), is superimposed onto this image. The pyridine nitrogen forms a water bridge to two neighboring isoleucines at the loop E domain (*8*, *13*). The three fluorine atoms of the compound, facing toward multiple directions, undergo various interactions with an interfacial region of loops C and D of the AChBP via hydrogen bonding and van der Waals contact. These interactions are fundamentally consistent with those observed in the insect nAChR structural model (*11*).

using SigmaPlot software (SPSS Inc., Chicago, IL). Insecticidal activity was evaluated with adult female houseflies via intrathoracic injection and topical application in the absence or the presence of a cytochrome P450 inhibitor [*O*-propyl *O*-(2-propynyl) phenylphosphonate], which serves as a synergist by reducing the oxidative detoxification rate (20).

RESULTS

Hydrophobicity and Photostability (Table 1). Six nitroimino (or cyanoimino) compounds (1–6) including IMI (1) and THIA (4) had log P_{ow} values of -0.16-1.26. In contrast, the 2-trifluor-oacetylimino [=NC(O)CF₃] analogues (7–16) showed the greatly enhanced log P_{ow} values (0.99–3.40). Acetylimino analogue [=NC(O)CH₃] (17) had a log P_{ow} of 1.32, which is significantly lower than that of the corresponding CF₃ compound (12) (3.06). Furthermore, the 2-trifluoroacetylmethylene [=CHC-(O)CF₃] compounds (18–22) retained a high level of hydrophobicity (log P_{ow} values of 0.80–2.10) relative to those of nitro or



Figure 2. Chemical structures of neonicotinoid analogues providing nitro (or cyano) and trifluoroacetyl substituents examined in the present investigation. The neonicotinoid compounds consist of three structural moieties.

Scheme 1. Preparation of Trifluoroacetyl Neonicotinoid Analogues



cyanoimino compounds. Among the =NC(O)CF₃ analogues, the hydrophobicity was varied by the heterocyclic moieties: that is, thiazolidine (12) or thiazoline (15) > imidazoline (8) > imidazolidine (7) > oxazolidine (11) or acyclics (10 and 16) > *N*-methylimidazolidine (9). In the series of 2-substituted thiazolidines [=NNO₂, =NC(O)CF₃, or =CHC(O)CF₃], their log P_{ow} values reduced in the order of chlorothiazolylmethyl (CTM) (5, 13, and 20) > chloropyridinylmethyl (CPM) (3, 12, and 19) > tetrahydrofurylmethyl (THFM) (6, 14, and 21).

On the ground level, sunlight of 290–400 nm wavelength affects the field stability of neonicotinoid insecticide (21, 22). Therefore, the λ_{max} is defined as a superficial index for neonicotinoid photostability and analogues with $\lambda_{max} \ge 290$ nm may possibly be photolabile. Nitro- or cyanoimino compounds (1–6) had λ_{max} values < 290 nm. Fascinatingly, the =NC(O)CF₃/CH₃ analogues (7–17) also showed the λ_{max} values < 290 nm, except for an analogue (15) with thiazoline heterocycle (307 nm). In marked contrast, all compounds with the =CHC(O)CF₃ substituent (18–22) had λ_{max} values > 290 nm. In addition, for a

Table 1. SARs of Neonicotinoid Insecticides with Nitro (or Cyano) and Trifluoroacetyl Pharmacophores

compound ^a				parameters		binding to Musca nAChR	toxicity to <i>Musca</i> LD_{50} (μ g/g female) ^c		
no.	Х	type	Y	log P _{ow}	λ_{max}	$IC_{50} (nM \pm SD) (n = 3)$	injection synergist ^d	topical synergist ^d	topical alone ^e
1	CPM	А	NNO ₂	0.57	269	10 ± 0.4	0.021	0.20	>100 (20%)
2	CPM	D	NNO ₂	-0.16	264	~100000 (53%) ^b	>1 (0%)	>10 (0%)	>100 (0%)
3	CPM	F	NNO ₂	1.16	279	24 ± 3.9	0.081	0.11	>100 (0%)
4	CPM	F	NCN	1.26	250	4.8 ± 0.3	0.032	0.14	>100 (30%)
5	CTM	F	NNO ₂	1.20	281	56 ± 1.8	0.11	0.60	>100 (0%)
6	THFM	F	NNO ₂	0.14	284	1310 ± 200	0.55	1.12	>100 (0%)
7	CPM	А	NC(O)CF ₃	1.92	248	68 ± 8.9	0.15	0.39	>100 (40%)
8	CPM	В	NC(O)CF ₃	2.41	270	22 ± 2.1	0.25	0.33	17.0
9	CPM	С	NC(O)CF ₃	0.99	247	50800 ± 8100	>1 (0%)	>10 (10%)	>100 (0%)
10	CPM	D	NC(0)CF ₃	1.53	244	>100000 (19%) ^b	>1 (0%)	>10 (0%)	>100 (0%)
11	CPM	Е	NC(O)CF ₃	1.56	254	2750 ± 110	>1 (0%)	>10 (20%)	>100 (20%)
12	CPM	F	NC(O)CF ₃	3.06	265	28 ± 1.5	0.060	0.040	2.0
13	CTM	F	NC(O)CF ₃	3.40	266	300 ± 51	0.60	0.51	>100 (0%)
14	THFM	F	NC(O)CF ₃	1.51	264	36600 ± 6400	>1 (6.7%)	4.0	>100 (0%)
15	CPM	G	NC(O)CF ₃	2.93	307	5.2 ± 0.4	0.020	0.043	1.1
16	CPM	Н	NC(O)CF ₃	1.70	263	44600 ± 2300	>1 (0%)	>10 (0%)	>100 (0%)
17	CPM	F	NC(O)CH ₃	1.32	262	1470 ± 170	>1 (0%)	3.2	80
18	CPM	А	CHC(O)CF ₃	1.76	293	3.0 ± 0.3	0.012	0.058	3.8
19	CPM	F	CHC(O)CF ₃	1.84	322	3.2 ± 0.4	0.0052	0.0040	0.75
20	CTM	F	CHC(O)CF ₃	2.10	321	78 ± 19	0.11	0.12	>100 (20%)
21	THFM	F	CHC(O)CF ₃	0.80	321	19200 ± 5000	>1 (3.4%)	>10 (10%)	>100 (0%)
22	CPM	G	CHC(0)CF ₃	1.83	352	1.2 ± 0.1	0.0038	0.0070	1.6
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^a Chemical structures are given in **Figure 2**. Compounds **1** and **4** are IMI and THIA, respectively. ^b Inhibition % at the indicated concentration. ^c Numbers in parentheses are mortality % at the indicated dose. ^d Pretreated with *O*-propyl *O*-(2-propynyl) phenylphosphonate at 100 µg/g female. ^e Chlorpyrifos as the control insecticide had a LD₅₀ value of 1.9 µg/g female as compared with those of other standard insecticides parathion, propoxur, dieldrin, and DDT of 1.3, 23, 0.7, and 14 µg/g female (*11, 23*).



Figure 3. Correlation plot between IC₅₀ and LD₅₀ values as indexes for receptor potency and intrinsic insecticidal activity (determined by intrathoracic injection with synergist pretreatment), respectively, of 14 neonicotinoid analogues. Eight inactive compounds showing LD₅₀ values of >1 μ g/g and very low binding affinities were excluded.

representative compound **12**, almost none of the hydrolytic product was detected in neutral aqueous solution until 96 h (data not shown).

Binding Affinity (Table 1). Nitroimino or cyanoimino analogues with imidazolidine and thiazolidine rings (1 and 3-5) had relatively high binding affinities, while the acyclic analogue was inactive. Among the 2-nitroiminothiazolidines, the binding potency was diminished depending on the substituent structure with the order of CPM (3) > CTM (5) > THFM (6). Trifluoroacetylimino analogues with imidazolidine (7), imidazoline (8), thiazolidine (12), and thiazoline (15) showed high binding affinities comparable to those of nitroimino neonicotinoids. The thiazoline analogue (15) displayed the highest affinity in the $=NC(O)CF_3$ series. However, the compounds with *N*-methylimidazolidine (9), oxazolidine (11), and two acyclic analogues (10 and 16) had largely reduced potencies. As with nitroimino compounds, the potency rank order was CPM (12) > CTM (13) > THFM (14) among the 2-trifluoroacetyliminothiazolidines. The =NC(O)CH₃ compound (17) was 53-fold less active than that of the corresponding CF_3 analogue (12). Intriguingly, the =CHC(O)CF₃ analogues with a CPM moiety (18, 19, and 22) were outstandingly active, although their CTM (20) and THFM (21) analogues had moderate and low binding affinities, respectively. The thiazoline compound (22) gave a higher binding potency than that of the thiazolidine analogue (19).

Insecticidal Activity (Table 1). Adult female houseflies were used for quantitative evaluation of insecticidal activity via two administration routes. First, intrathoracic injection of test compounds into flies, which were pretreated with a cytochrome P450 inhibitor (synergist), was determined to define intrinsic toxicity. Second, test compounds were administrated by a topical application route (in the presence or the absence of synergist), involving the penetration aspect of a compound into the insect body through the epidermal cuticle.

Importantly, the intrinsic toxicities of 14 neonicotinoids were well-correlated to their target site potencies (r = 0.92) (Figure 3). Eight inactive compounds had very low binding affinities as well. The toxicity range of =NC(O)CF₃ analogues was similar to that of nitroimino (or cyanoimino) neonicotinoids. The =CHC-(O)CF₃ compounds (18, 19, and 22) exhibited excellent intrinsic toxicity, except for compounds 20 and 21 with CTM and THFM moieties. Intrinsically, toxic compounds were generally insecticidal via topical application experiment in the presence of a synergist. Most of test compounds including IMI (1) and THIA (4) were inactive in the topical application without the synergist pretreatment. In marked contrast, six compounds with =NC-(O)CF₃ and =CHC(O)CF₃ pharmacophores (8, 12, 15, 18, 19, (0)and 22) provocatively showed outstanding insecticidal activity, rivaling those of an organophosphate insecticide chlorpyrifos and other standard insecticides (Table 1 footnote) (11, 23).

DISCUSSION

Exploration of novel nicotinic pharmacophore chemotypes, undergoing the unique or atypical binding mechanism(s), may expand the insecticidal spectrum and circumvent the possible resistance caused by activated detoxification systems and a modified target site (24, 25). The present investigation considers exhaustive SARs of neonicotinoid analogues with a trifluoroacetyl pharmacophore, which may advance the chemical property and insecticidal effectiveness of the present nitro and cyano neonicotinoids.

The SARs of trifluoroacetyl neonicotinoids, in terms of target site potency and toxicity, are nearly consistent with those of nitro or cyano neonicotinoid analogues (20, 26-28). Interestingly, comparative SARs between the $=NC(O)CF_3$ and the =CHC- $(O)CF_3$ analogues are almost identical to those between the nitroimino (=NNO₂) and nitromethylene (=CHNO₂) neonicotinoids. The receptor binding affinity is predictive of the intrinsic toxicity based on a good correlation. A specific combination of three structural moieties determines the potency at the target site. The $=NC(O)CH_3$ is less active than the corresponding =NC-(O)CF₃ compound, establishing that the trifluoromethyl head plays an important role on the selective binding interaction with the receptor subsite(s) via H-bonding and van der Waals contact (11, 12). Furthermore, an electron-withdrawing effect of the CF_3 apparently facilitates π -stacking interaction of the amidine moiety with the loop C aromatic residue (11, 14). The neonicotinoid =NC(O)CF₃ substituent is embraced by an interfacial niche between the α and the β subunits, whereas the nitroguanidine or cyanoamidine moiety of IMI or THIA is primarily recognized by the α subunit (9, 11).

The neonicotinoids generally have low log Pow values (-0.7-1.3), which makes the basis of their outstanding plant systemic performance to control the piercing-sucking insects (3). Some organophosphates (1-5.5) and methylcarbamates (-1-3)also have good plant-mobile action. However, the more lipophilic organochlorines (5.5-7.5) and pyrethroids (4-9) are efficacious against *lepidopterous* larvae as contact insecticides (3). The =NC-(O)CF₃ and =CHC(O)CF₃ substituents (log P_{ow} values 1.0-3.4 and 0.8-2.1, respectively) greatly enhance the hydrophobicity of nitro- and cyanoimino neonicotinoids. On the basis of the present findings, the increased hydrophobicity of trifluoroacetyl analogues presumably improves the penetrability of the neonicotinoid compound into the insect integument, thereby conferring the enhanced insecticidal efficacy relative to that of IMI (1) or THIA (4). Moreover, the neonicotinoid with $=NC(O)CF_3$ pharmacophore appears to retain adequate photostability comparable to that of the =NNO₂ or =NCN neonicotinoid; yet, the =CHC-(O)CF₃ analogues are conceivably photolabile as with the =CHNO₂ compounds (21, 22).

In summary, the present SAR investigation identifies several trifluoroacetyl neonicotinoids with excellent insecticidal effectiveness associated with the high receptor potency and the enhanced hydrophobicity, thereby illustrating that the nicotinic receptor target further warrants continuing study to discover novel nicotinic insecticides with unique biological properties.

ABBREVIATIONS USED

AChBP, acetylcholine binding protein; IMI or [³H]IMI, imidacloprid or its tritiated radioligand; log P_{ow} , partition coefficient of compound between 1-octanol and water; nAChR, nicotinic acetylcholine receptor; SAR, structure–activity relationship; THIA, thiacloprid.

Supporting Information Available: Synthesis procedures and analysis data for new products. This material is available free of charge via the Internet at http://pubs.acs.org.

LITERATURE CITED

 Kagabu, S. Molecular design of neonicotinoids: Past, present, and future. In *Chemistry of Crop Protection*; Voss, A., Ramos, G., Eds.; Wiley-VCH: Weinheim, 2003; pp 193–212.

- (2) Tomizawa, M.; Casida, J. E. Selective toxicity of neonicotinoids attributable to specificity of insect and mammalian nicotinic receptors. *Annu. Rev. Entomol.* 2003, 48, 339–364.
- (3) Tomizawa, M.; Casida, J. E. Neonicotinoid insecticide toxicology: Mechanisms of selective action. *Annu. Rev. Pharmacol. Toxicol.* 2005, 45, 247–268.
- (4) Matsuda, K.; Shimomura, M.; Ihara, M.; Akamatsu, M.; Sattelle, D. B. Neonicotinoids show selective and diverse actions on their nicotinic receptor targets: Electrophysiology, molecular biology, and receptor modeling studies. *Biosci., Biotechnol., Biochem.* 2005, 69, 1442–1452.
- (5) Jeschke, P.; Nauen, R. Neonicotinoids—From zero to hero in insecticide chemistry. *Pest Manage. Sci.* 2008, 64, 1084–1098.
- (6) Tomizawa, M.; Talley, T. T.; Maltby, D.; Durkin, K. A.; Medzihradszky, K. F.; Burlingame, A. L.; Taylor, P.; Casida, J. E. Mapping the elusive neonicotinoid binding site. *Proc. Natl. Acad. Sci. U.S.A.* 2007, 104, 9075–9080.
- (7) Tomizawa, M.; Maltby, D.; Talley, T. T.; Durkin, K. A.; Medzihradszky, K. F.; Burlingame, A. L.; Taylor, P.; Casida, J. E. Atypical nicotinic agonist bound conformations conferring subtype selectivity. *Proc. Natl. Acad. Sci. U.S.A.* **2008**, *105*, 1728–1732.
- (8) Talley, T. T.; Harel, M.; Hibbs, R. H.; Radić, Z.; Tomizawa, M.; Casida, J. E.; Taylor, P. Atomic interactions of neonicotinoid agonists with AChBP: Molecular recognition of the distinctive electronegative pharmacophore. *Proc. Natl. Acad. Sci. U.S.A.* 2008, 105, 7606–7611.
- (9) Tomizawa, M.; Casida, J. E. Molecular recognition of neonicotinoid insecticides: the determinants of life or death. Acc. Chem. Res. 2009, 42, 260–269.
- (10) Tomizawa, M.; Talley, T. T.; Park, J. F.; Maltby, D.; Medzihradszky, K. F.; Durkin, K. A.; Cornejo-Bravo, J. M.; Burlingame, A. L.; Casida, J. E.; Taylor, P. Nicotinic agonist binding site mapped by methionine- and tyrosine-scanning coupled with azidochloropyridinyl photoaffinity labeling. J. Med. Chem. 2009, 52, 3735–3741.
- (11) Tomizawa, M.; Kagabu, S.; Ohno, I.; Durkin, K. A.; Casida, J. E. Potency and selectivity of trifluoroacetylimino and pyrazinoylimino nicotinic insecticides and their fit at a unique binding site niche. *J. Med. Chem.* **2008**, *51*, 4213–4218.
- (12) Ohno, I.; Tomizawa, M.; Durkin, K. A.; Naruse, Y.; Casida, J. E.; Kagabu, S. Molecular features of neonicotinoid pharmacophore variants interacting with the insect nicotinic receptor. *Chem. Res. Toxicol.* **2009**, *22*, 476–482.
- (13) Ohno, I.; Tomizawa, M.; Durkin, A.; Casida, J. E.; Kagabu, S. Neonicotinoid substituents forming a water bridge at the nicotinic acetylcholine receptor. J. Agric. Food Chem. 2009, 57, 2436–2440.
- (14) Tomizawa, M.; Zhang, N.; Durkin, K. A.; Olmstead, M. M.; Casida, J. E. The neonicotinoid electronegative pharmacophore plays the crucial role in the high affinity and selectivity for the *Drosophila* nicotinic receptor: An anomaly for the nicotinoid cation-π interaction model. *Biochemistry* 2003, *42*, 7819–7827.
- (15) Wolf, H.; Abbink, J.; Becker, B.; Homeyer, B.; Stendel, W.; Moriya, K. Preparation and testing of (heteroarylalkyl)-substituted 5- and 6-membered heterocycles as insecticides and ectoparasiticides. Ger. Offen., DE3639877, 1988.
- (16) Shiokawa, K.; Tsuboi, S.; Moriya, K.; Hattori, Y.; Murata, S.; Shibuya, K. Preparation of pyridine and thiazole compounds containing trifluoroacetyl group as insecticides. Jpn. Kokai Tokkyo Koho, JP03220176, 1991.
- (17) Aoki, I.; Minamida, I. Preparation of nitroguanidines as agrochemicals. Jpn. Kokai Tokkyo Koho, JP05112521, 1993.
- (18) Kagabu, S.; Ishihara, R.; Hieda, Y.; Nishimura, K.; Naruse, Y. Insecticidal and neuroblocking potencies of variants of the imidazolidine moiety of imidacloprid-related neonicotinoids and the relationship to partition coefficient and charge density on the pharmacophore. J. Agric. Food Chem. 2007, 55, 812–818.
- (19) Tomizawa, M.; Latli, B.; Casida, J. E. Novel neonicotinoid-agarose affinity column for *Drosophila* and *Musca* nicotinic acetylcholine receptors. J. Neurochem. **1996**, 67, 1669–1676.
- (20) Liu, M.-Y.; Lanford, J.; Casida, J. E. Relevance of [³H]imidacloprid binding site in house fly head acetylcholine receptor to insecticidal

activity of 2-nitromethylene- and 2-nitroimino-imidazolidines. *Pestic. Biochem. Physiol.* **1993**, *46*, 200–206.

- (21) Kagabu, S.; Medej, S. Stability comparison of imidacloprid and related compounds under simulated sunlight, hydrolysis conditions, and to oxygen. *Biosci., Biotechnol., Biochem.* 1995, 59, 980–985.
- (22) Kagabu, S.; Akagi, T. Quantum chemical consideration of photostability of imidacloprid and related compounds. *J. Pestic. Sci.* **1997**, *22*, 84–89.
- (23) Palmer, C. J.; Casida, J. E. 1-(4-Ethynylphenyl)-2,6,7-trioxabicyclo-[2.2.2]octanes: A new order of potency for insecticides acting at the GABA-gated chloride channel. J. Agric. Food Chem. 1989, 37, 213–216.
- (24) Karunker, I.; Morou, E.; Nikou, D.; Nauen, R.; Sertchook, R.; Stevenson, B. J.; Paine, M. J. I.; Morin, S.; Vontas, J. Structural model and functional characterization of the *Bemisia tabaci* CYP6CM1vQ, a cytochrome P450 associated with high levels of imidacloprid resistance. *Insect Biochem. Mol. Biol.* 2009, *39*, 697– 706.

- (25) Liu, Z.; Williamson, M. S.; Lansdell, S. J.; Han, Z.; Denholm, I.; Millar, N. S. A nicotinic acetylcholine receptor mutation (Y151S) causes reduced agonist potency to a range of neonicotinoid insecticides. J. Neurochem. 2006, 99, 1273–1281.
- (26) Tomizawa, M.; Yamamoto, I. Structure-activity relationships of nicotinoids and imidacloprid analogs. J. Pestic. Sci. 1993, 18, 91–98.
- (27) Yamamoto, I.; Tomizawa, M.; Saito, T.; Miyamoto, T.; Walcott, E. C.; Sumikawa, K. Structural factors contributing to insecticidal and selective actions of neonicotinoids. *Arch. Insect Biochem. Physiol.* **1998**, *37*, 24–32.
- (28) Honda, H.; Tomizawa, M.; Casida, J. E. Insect nicotinic acetylcholine receptors: Neonicotinoid binding site specificity is usually but not always conserved with varied substituents and species. J. Agric. Food Chem. 2006, 54, 3365–3371.

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